



Transistors à nanofils de silicium pour la détection de ions et protéines



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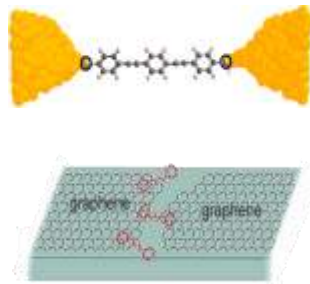
***Since Oct. 1st, 2016, also at
Swiss Federal Laboratories for
Materials Science & Technology
Dübendorf, Switzerland***

research activities

nanoscale electronics & molecular junctions

Individual molecular junctions

A. Vladyka, J. Overbeck,
M. El Abbassi, et al.



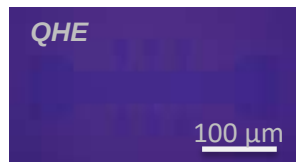
Networks of molecular junctions & hybrid devices

A. Vladyka et al.



Graphene & metrology

K. Thodkar, M. El abbassi,
A. Vladyka, J. Overbeck et al.



Optoelectronics & plasmonics

J. Overberck, T. Fröhlich et al.

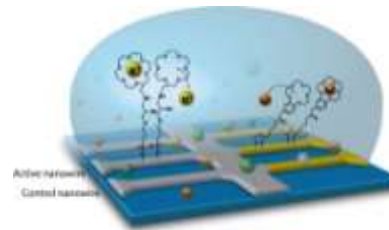
ions and biochemicals detection

Si nanowires FETs

ions
bio-molecules

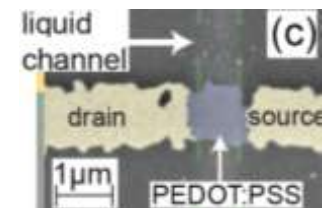
pathogens

pH, Na⁺, K⁺, Ca²⁺, F⁻
lectin (FimH)
sugar binding protein
Avails Medical



M. Baghernejad, A. Fanget,
O. Synhaivska,
R. Stoop, M. Wipf et al.

OEECTs



R. Stoop, M. Sessolo,
H. Bolink (Valencia) et al.

Silicon-based transistors for sensing

motivation: diagnostics & drug screening

- *specific detection of biochemicals at low cost*
- *time-resolved, label-free, quantitative, portable*
- *statistically relevant datasets for multiple analytes*

⇒ **simplify point of care-, point of prescription- & possibly home-diagnostics**



see e.g. Erickson et al., *Lab on a chip* (2014)

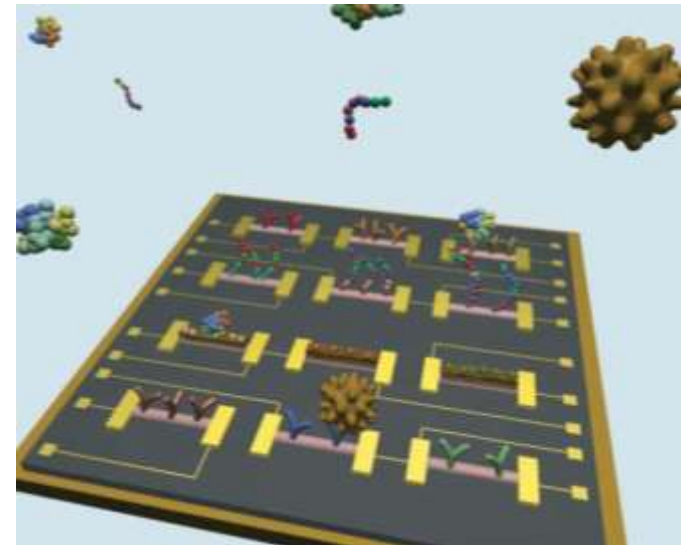
Public Health, Nanotechnology & Mobility program (PHeNoM, Cornell)

Silicon-based biochemical sensors

- high-integration
- industrial fabrication processes

⇒ ***Si nanoribbons* operated as *ion-sensitive field-effect transistors (ISFETs)***

transducing biochemical reactions in electrical signals



F. Patolsky, C.M. Lieber et al. MRS Bulletin 2007

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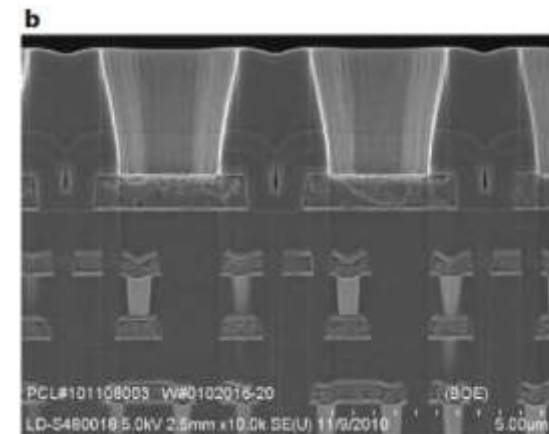
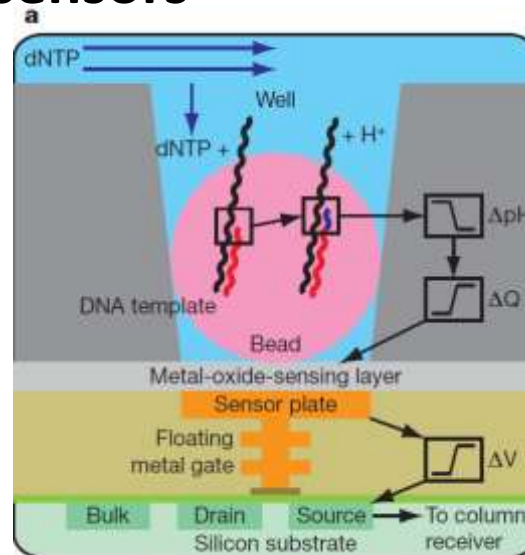
see e.g. Erickson et al., *Lab on a chip* (2014)

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Silicon-based biochemical sensors

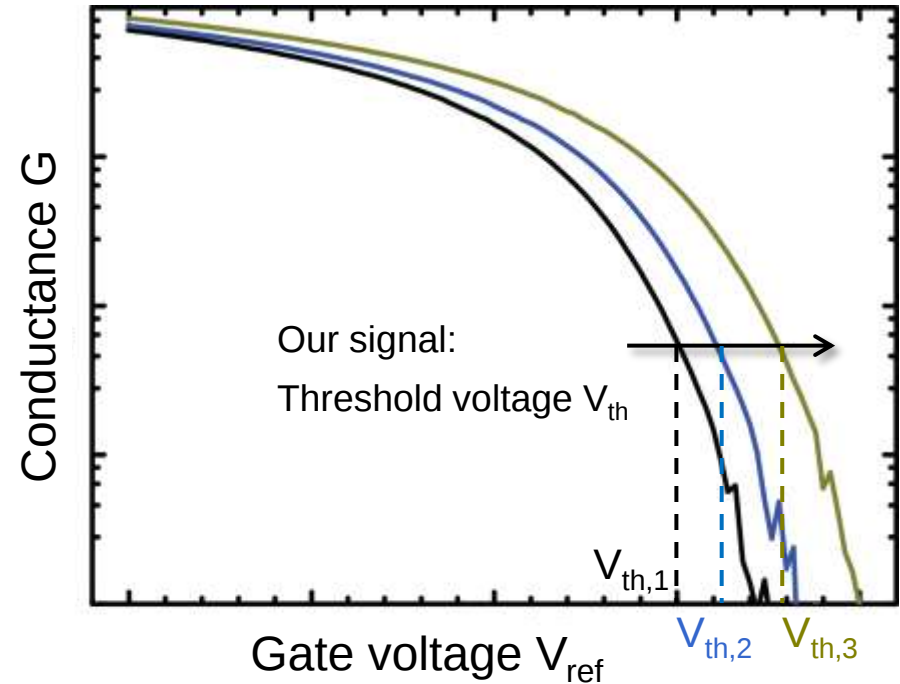
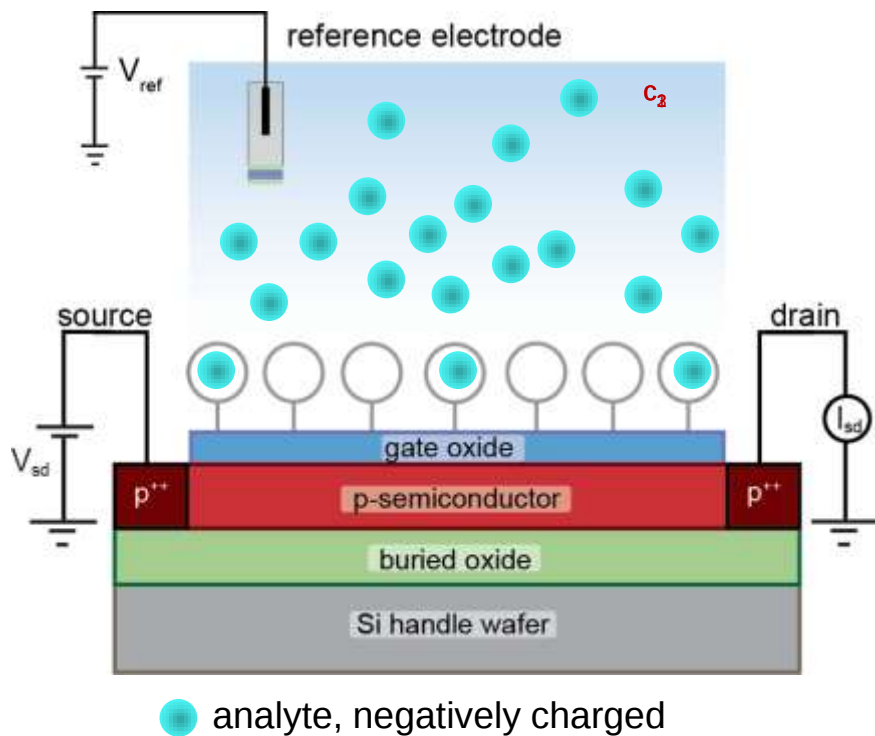
- high-integration
- industrial fabrication processes

example: non-optical, on-chip sequencing using integrated ISFETs for pH monitoring



Rothberg et al., *Nature* (2011)
Ion-Torrent technology

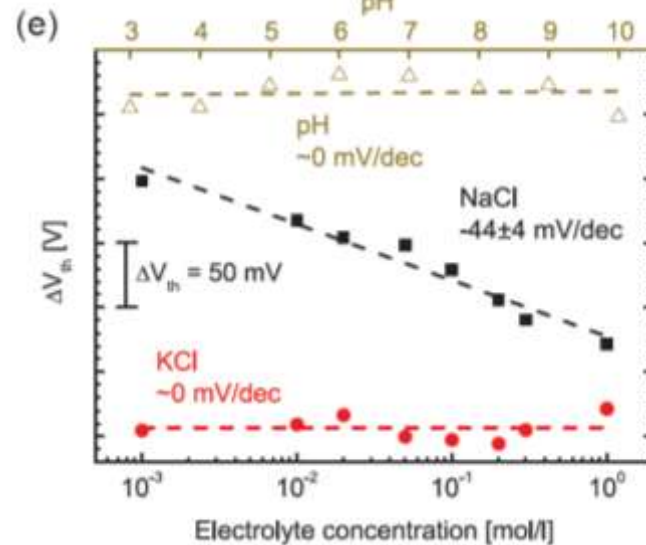
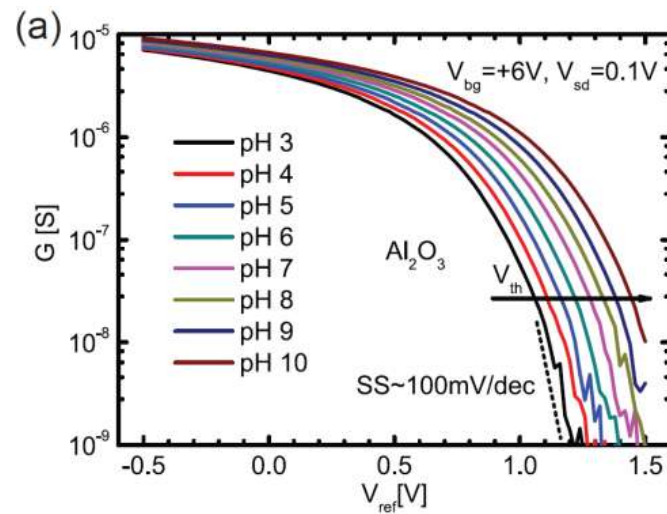
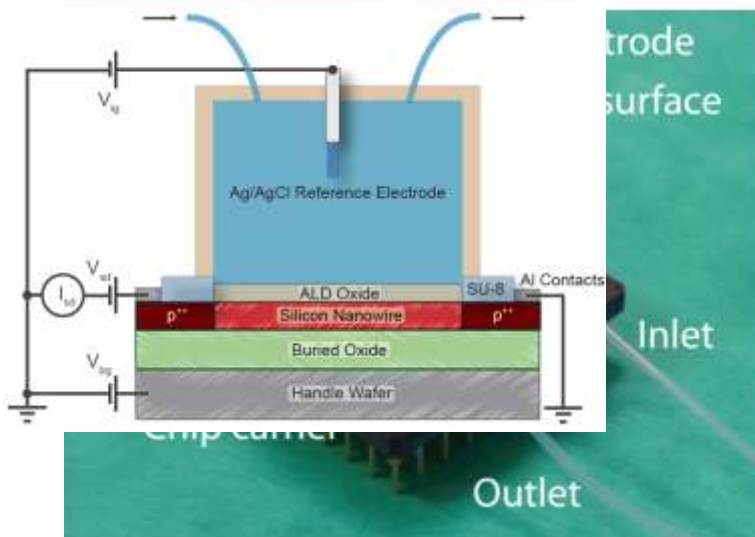
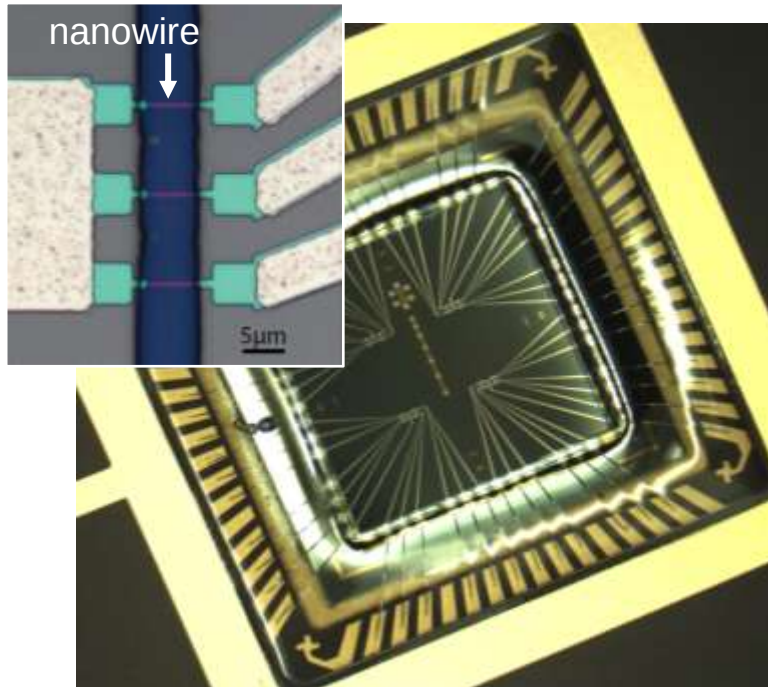
ISFETs as potentiometric biochemical sensors



Theoretical limit: Nernst response

$$\Delta V_{th} = V_{th,2} - V_{th,1} = 2.3 \frac{kT}{e} \log_{10} \left(\frac{c_2}{c_1} \right) \approx 59.6 \text{ mV} \cdot \log_{10} \left(\frac{c_2}{c_1} \right)$$

ions sensing with Si FETs

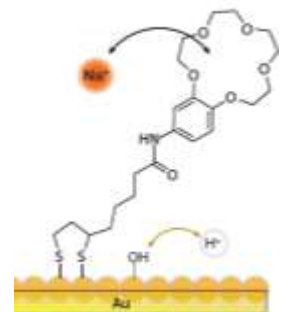


pH sensing

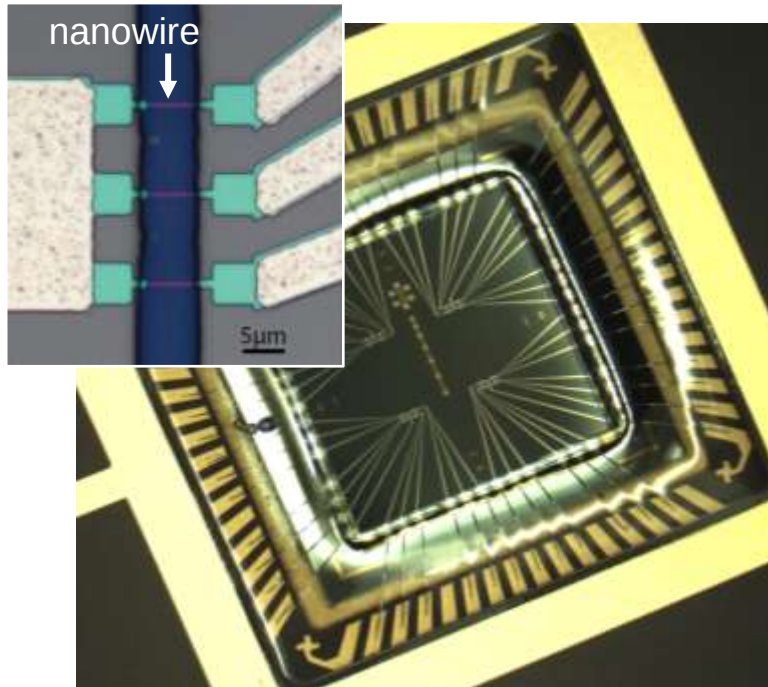
protonation / deprotonation
of surface oxide groups

oxide	solution
Al_2O_3	OH_2^+
or	OH
HfO_2	O^-

ion sensing functionalization



ions sensing with Si FETs



- pH & surface passivation
- ions Na^+ , K^+ , Ca^{2+} , F^-
HfO₂, Al₂O₃ or Au surfaces
- noise, size, s/n ratio, competition effects
- **quantifying protein-ligand interactions**
state of the art: surface plasmon resonance (SPR)
limited throughput, cost-intensive

⇒ **can SiNW do the job ?**

*to date mostly DNA and biotin-streptavidin interactions,
see e.g. Reed et al., Nat. Nano (2012)*

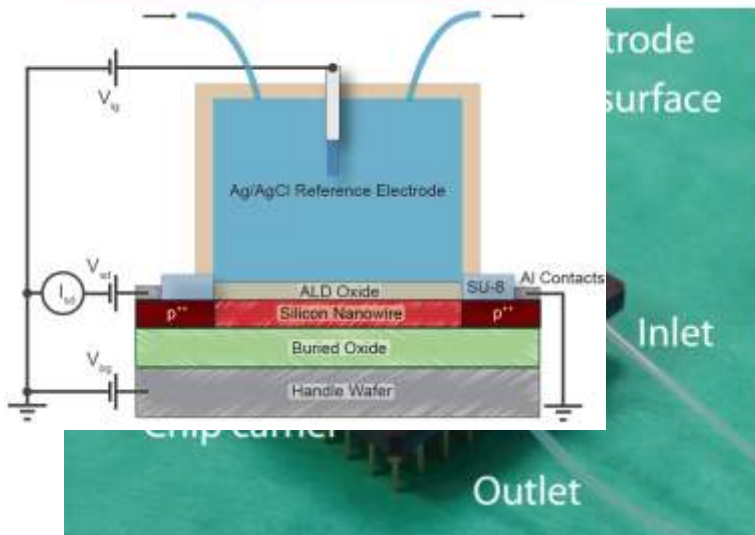
Test system: FimH (*bacterial lectin*)

with B. Ernst, G. Navarra, Dpt. Pharmacology, Uni Basel

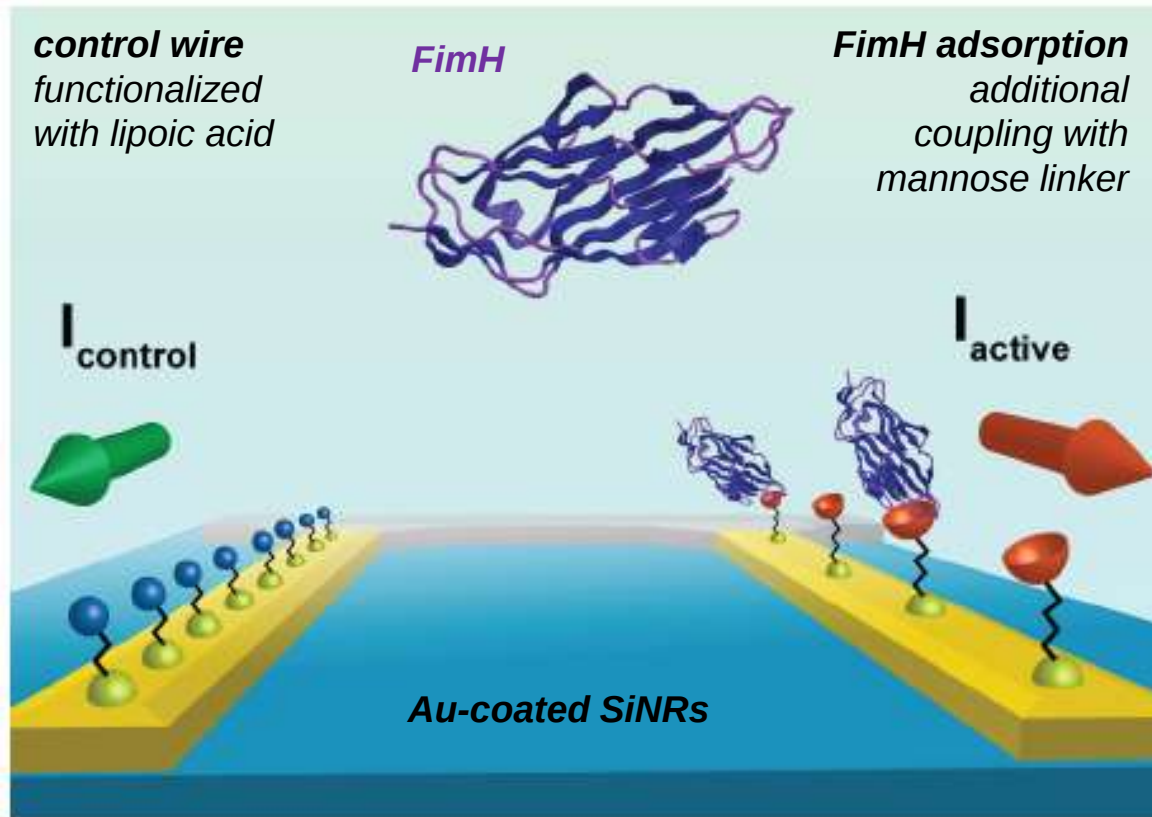
UTI therapy: high-affinity FimH antagonists

⇒ affinity screening tests: **SPR**

Ernst et al., J. Med. Chem. 2010, 2012; Chemmedchem 2012

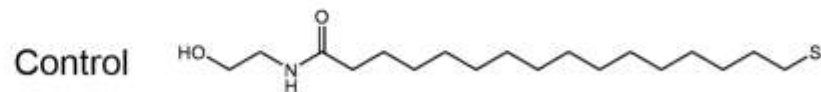


FimH detection kinetics

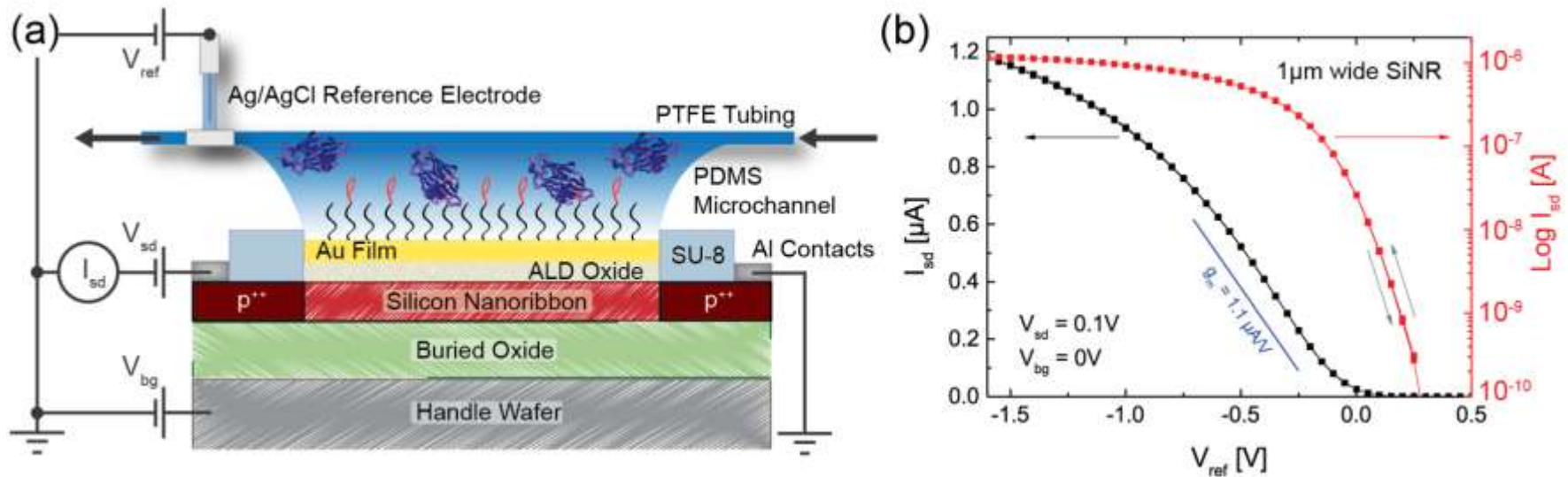


functionalization

- mercaptohexadecanoic acid (MHDA)
- amine coupling for ligand



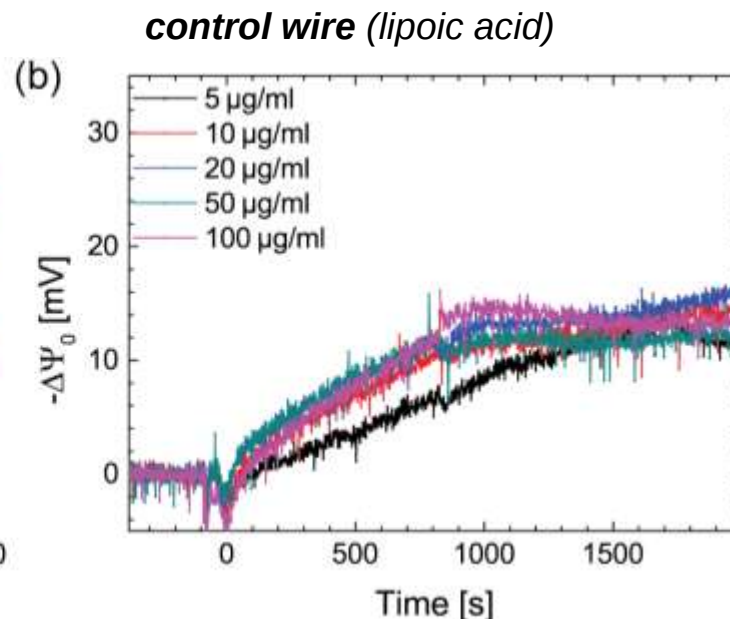
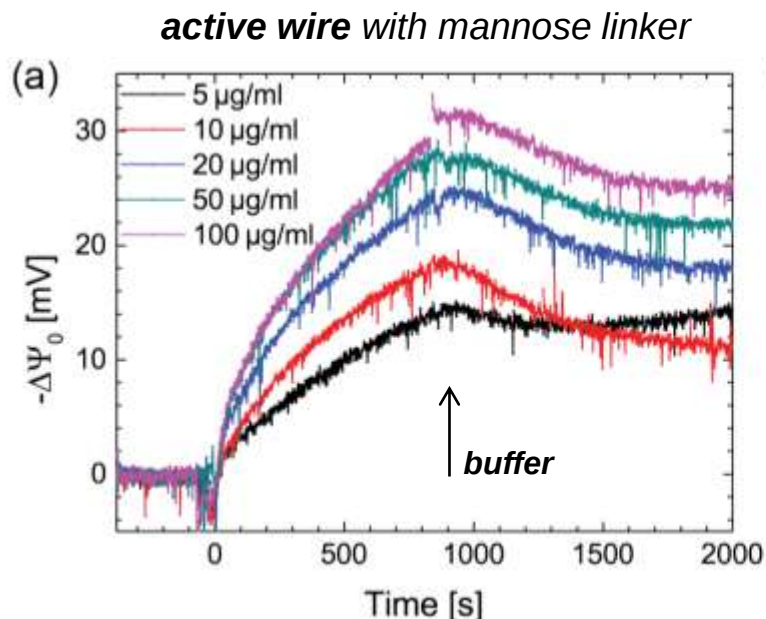
beyond ions



- reduced ionic strength buffer: 10mM HEPES, pH 8 (Debye length $\lambda_D \geq 3$ nm)
ensure that the proteins are within the electrical double to affect the surface potential
- at pH 8: FimH neg. charged $\Rightarrow I_{sd}$ increase upon binding
- SiNW operated in **linear region** (constant transconductance g_m)

FimH binding kinetics vs concentration

SiNRs



SiNRs

- **non-specific adsorption** (control wire)
lipophilic character of MHDA layer
- **signal only a function of surf. pot. change**
site binding model see Reed et al., Nat. Nano 2012.

surface bound ligands/m²

charge per analyte

$$\Delta\Psi_0 = -\Delta V_{th} = -\frac{\Delta I_{sd}}{g_m} = \frac{q_A}{C_0} [B]_0 \times \frac{[A]}{K_D + [A]}$$

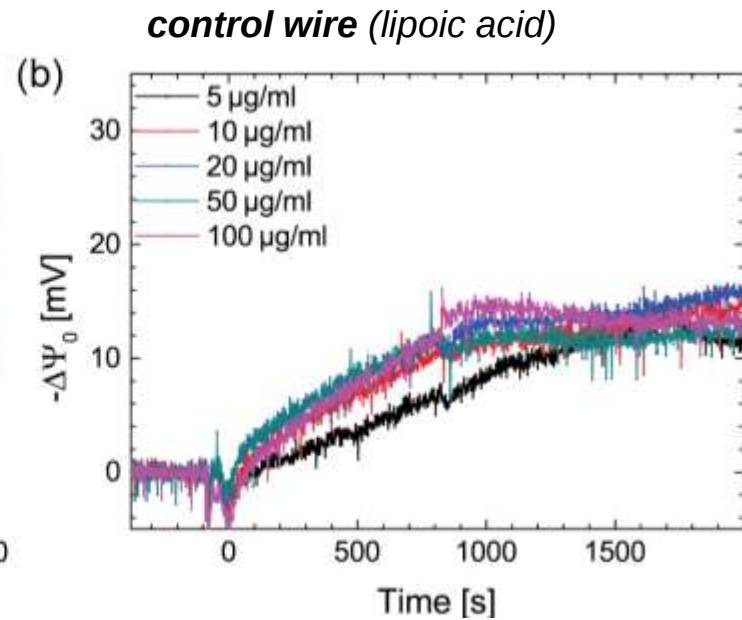
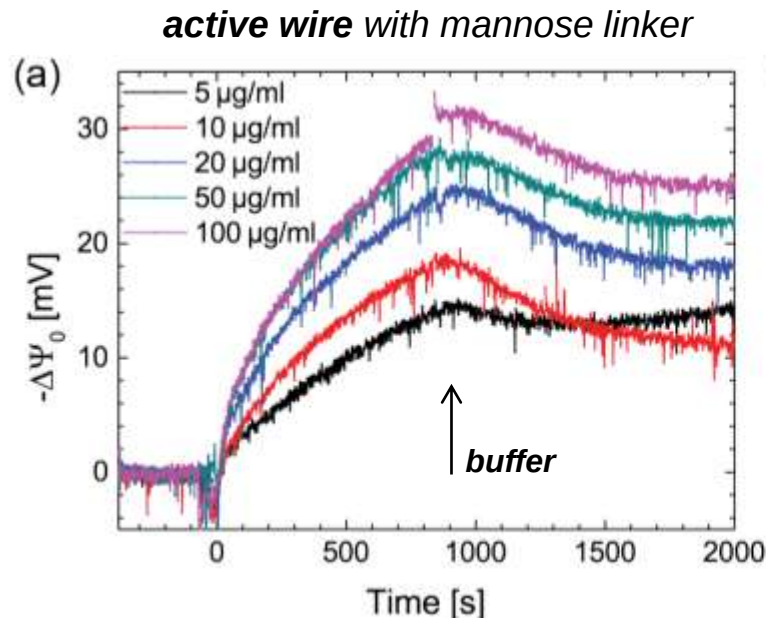
$$(g_m = \partial I_{sd} / \partial V_{ref})$$

equilibrium
dissociation
constant

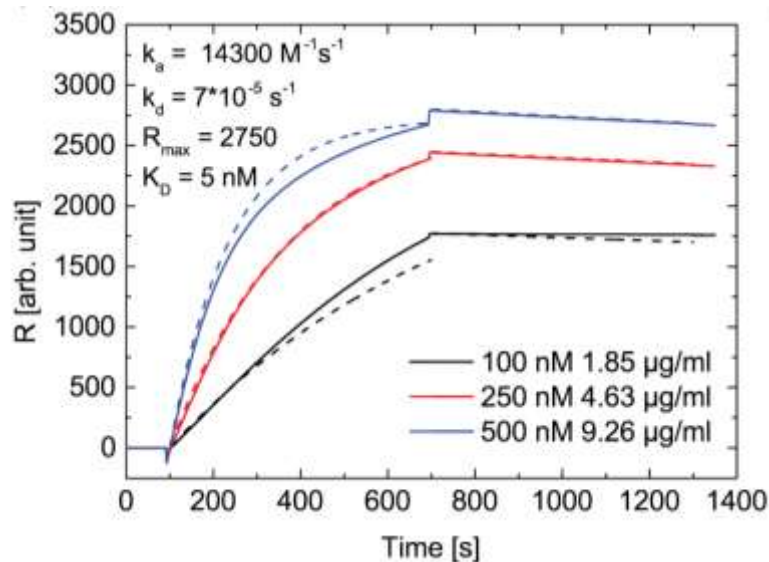
analyte bulk
concentration

FimH binding kinetics vs concentration

SiNRs



SPR



SiNRs

- **non-specific adsorption** (control wire)
lipophilic character of MHDA layer
- **signal only a function of surf. pot. change**
see Reed et al., Nat. Nano 2012.

SPR (Biacore T200, Navarra, Ernst et al., Basel)

- **saturation at lower FimH concentrations**
- **dissociation less pronounced**
dashed lines: Langmuir kinetics, $K_D \sim 5\text{nM}$ is obtained.

FimH binding kinetics vs concentration

comparison SiNRs & SPR

- different **association and dissociation rates** (k_a , k_d)

NB: variations between SPR systems!

Cannon et al., *Anal. Biochem.* 2004; Katsamba et al., *Anal. Biochem.* 2006

possible origins of differences

- flow rate at sensor surface & different surface areas**

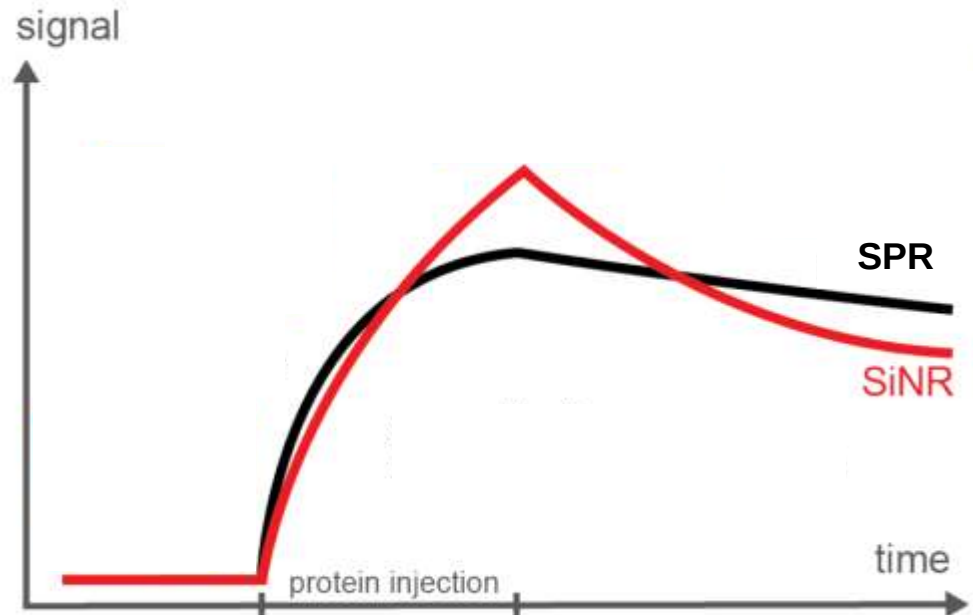
- *FimH-mediated bacterial **adhesion** affected by shear forces*
see e.g. Vogel et al., *J. Bacteriol.* 2007, *J. Biol. Chem.* 2008

- **re-adsorption** of proteins in flow

- different effective protein concentration (fluidics)**

- different sensing mechanisms: optical** ($\lambda_{\text{evan}} \sim 300\text{nm}$) **or charge** ($\lambda_D \sim 3\text{nm}$)
⇒ protein surface rearrangements affect SiNRs stronger than SPR, **longer time const.**

Rabe et al., *Adv. Colloid Interface Sci.* 2011, Roach et al., *JACS* 2005



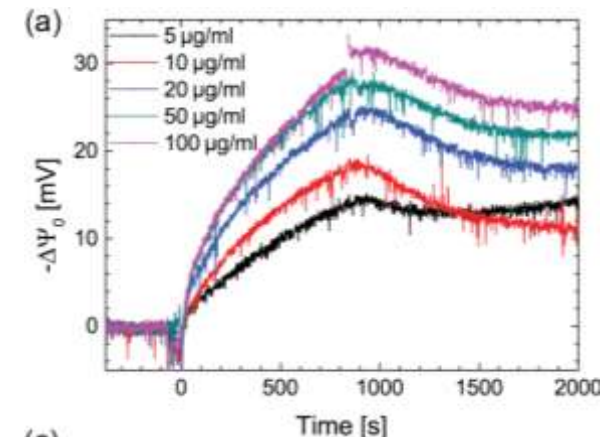
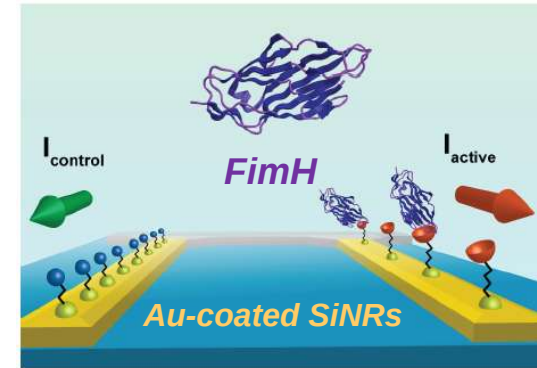
Fluidic channel	BioFET	Biacore
Flow rate	26 $\mu\text{L}/\text{min}$	20 $\mu\text{L}/\text{min}$
Height	100 μm	40 μm
Width	500 μm	500 μm
Length	4 mm	2.4 mm
Volume	$\approx 0.2 \mu\text{L}$	$\approx 0.05 \mu\text{L}$

conclusion & outlook

Si nanoribbon transistors as biosensors

- **time-resolved & label-free** detection of FimH with s/n ratio >700
- **Au surface** for strongly reduced pH response & suitable surface chemistry for direct comparison with SPR systems
- quantitative detection by taking into account **competing reactions**
- enhanced **sensitivity to surface rearrangements** as compared to SPR

⇒ *ISFETs for diagnosis & drug discovery*



Outlook: transmembrane proteins functionality & bacterial activity

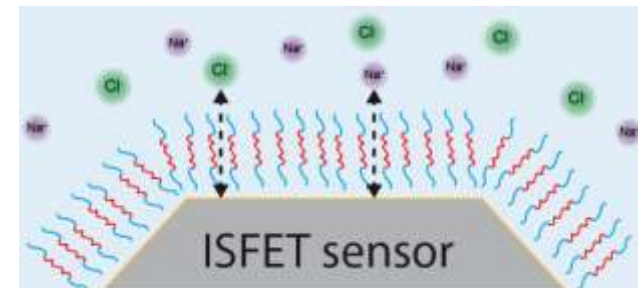
M. Baghernejad, A. Fanget et al.,
with C. Palivan, W. Meier et al., (Basel),
D. Fotiadis et al. (Bern)
A. Jesorka et al. (Chalmers)



NCCR
Molecular Systems
Engineering



EU H2020
Record It (FET)



start-up collaboration



"Accurate, rapid, digital detection of infections in any bodily fluid to indicate in real-time which drugs will be most effective at the point-of-prescription"

<http://www.availsmedical.com/>
Menlo Park (CA) USA



O. Knopfmacher
(CEO)



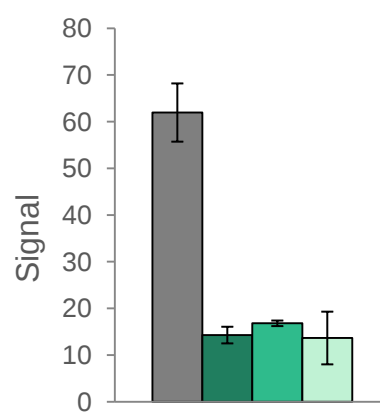
M. Herget
(CTO)



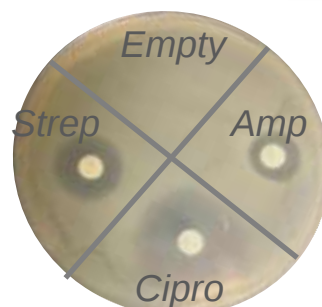
Axel Fanget

Avails AST technology vs disk diffusion to monitor microorganisms resistance to anti-biotics

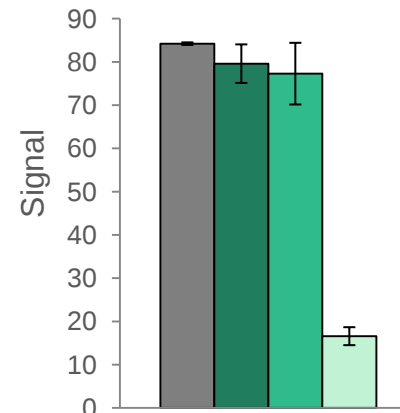
large signal: resistant



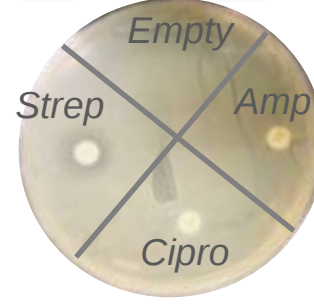
Empty Amp



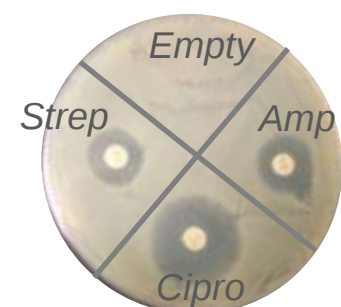
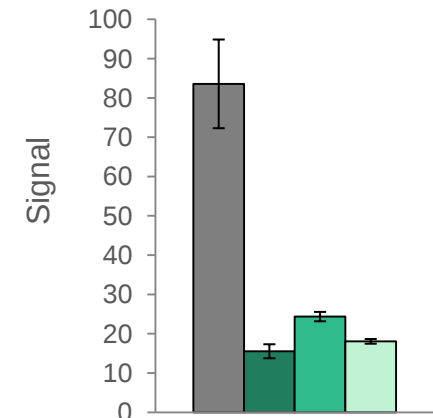
E.Coli



Cipro Strep



Clinical Sample 15



Clinical Sample 24

Acknowledgment

- Maria El Abbassi
- Kishan Thodkar
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- Axel Fanget
- Yves Mermoud

Former coworkers

R. Stoop (Barcelona), **M. Wipf** (Yale Univ.), A. Tarasov (BioMedX, Heidelberg), W. Fu (Jülich).

Nanoelectronics, Uni Basel C. Schönenberger

Collaborations

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O. Knopmacher, M. Herget (Avails Medical, USA)
E. Constable, C. Housecroft et al. (Basel)
G. Navarra, B. Ernst (Basel)
C. Palivan, W. Meier et al. (Basel)
D. Vuillaume et al. (IEMN, Lille)
A. Jesorka et al. (Chalmers)



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- European Union FP7 ITN & Strep and H2020 FET programs
- SNSF NCCR Molecular Systems Engineering
- Avails Medical, USA
- nanotera SNSF



EU FP7 & H2020



MOLESCO (ITN)
Record It (FET)

avails
MEDICAL

